

**AMENDMENTS TO THE SPECIFICATION**

**Please replace paragraph [15] of the originally filed specification with the following:**

FIG. 3 depicts comparisons of Fragment C, recombinant Fragment C and the Tetanus Toxoid with Group C Meningococcal Polysaccharide (GCMP) conjugates.

**Please replace paragraph [17] of the originally filed specification with the following:**

FIG. 5 depicts type-specific IgG elicited by Group B Streptococcus polysaccharide (GBSP) conjugates with Tetanus Toxoid or recombinant Fragment C.

**Please replace paragraph [32] of the originally filed specification with the following:**

Preparation of Group C Meningococcal [[C]] Polysaccharides (GCMP), Group Y Meningococcal Polysaccharides (GYMP) and Group W Meningococcal Polysaccharides (GWMP)  
Polysaccharides:

**Please replace paragraph [36] of the originally filed specification with the following:**

Polysaccharide capture by ultrafiltration (UF) with a 300 kDa molecular weight cut off (MWCO) membrane:

**Please replace paragraph [37] of the originally filed specification with the following:**

Approximately 1 3 L of cell-free microfiltered fermentation permeate is concentrated by UF to approximately 1 liter using a Biomax 300K Pellicon PELLICON® membrane (0.5 m<sup>2</sup>). The

concentrated retentate is diafiltered 12× against 1M [[NaCl]]NaCl and then [[LOX]]10× against DI water. It is further concentrated to approximately 0.2 L and collected.

**Please replace paragraph [15] of the originally filed specification with the following:**

The 300K retentate solution (ca 5 mg PS/mL) was adjusted to a final concentration of 2N NaOH and placed in an oven set to 80° C. for 16-18 hrs. After the reaction mixture had cooled off to less than 50° C., it was diluted into 10 L of DI water. After concentration through a 30 kDa MWCO Pellicon PELLICON® membrane, the concentrated retentate was diafiltered 12 times against 1 M NaCl and then 10 times against DI water. It was further concentrated to approximately 0.2 L and collected.

**Please replace paragraph [41] of the originally filed specification with the following:**

The retentate solution was transferred to a teflon reaction and sodium acetate (NaOAc) was added to a final concentration of 0.1 N. The reaction mixture was adjusted to pH5 using 6N HCl and placed in a water bath set to 7° C. It was shaken at 65 rpm until the polysaccharide reached a target MW of approximately 10-20 kDa as measured by Size Exclusion Chromatography Multi-Angle Laser Light Scatter (SEC-MALLS) using a Superose 12 (Pharmacia) column.

**Please replace paragraph [43] of the originally filed specification with the following:**

The pH of the solution was adjusted to 8 with 6N [[HCl]] HCl solution, and acetic anhydride was then added dropwise at room temperature to a final concentration of 0.8 M acetic anhydride. SN NaOH was used to keep the reaction mixture pH between 7 and 9. After completion of the

reaction, the pH of the reaction mixture was increased to 13, and the mixture stirred an additional 1.5 hr. The reaction pH was then adjusted to pH 8 with 6N [[HCl]]HCl solution. The reaction mixture was poured into 4 L of 1 M [[NaCl]]NaCl, concentrated to approximately 1 L using a Biomax 100K Pellicon PELLICON® membrane ( $0.5\text{ m}^2$ ) and the permeate collected. The 100K final permeate is concentrated by UF to approximately 1 liter using a Biomax 5K Pellicon PELLICON® membrane ( $0.5\text{ m}^2$ ). The concentrated retentate is diafiltered 10 times against DI water, then concentrated to approximately 0.2 L and collected. The fragmented polysaccharide was then activated with sodium metaperiodate to generate aldehyde groups in its sialic acid residues.